



# Rat ORM1 / alpha 1 Acid Glycoprotein ELISA Kit

Enzyme Immunoassay for the quantification of orosomuroid 1 in Plasma,  
Serum and Urine

Catalog number: ARG83480

Package: 96 wells

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For research use only. Not for use in diagnostic procedures.

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## **INTRODUCTION**

Functions as a transport protein in the blood stream. Binds various ligands in the interior of its beta-barrel domain. Also binds synthetic drugs and influences their distribution and availability in the body. Appears to function in modulating the activity of the immune system during the acute-phase reaction.

## **PRINCIPLE OF THE ASSAY**

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for ORM1 / alpha 1 Acid Glycoprotein has been pre-coated onto a microtiter plate. Standards or samples are pipetted into the wells and any ORM1 / alpha 1 Acid Glycoprotein present is bound by the immobilized antibody. After washing away any unbound substances, a Horseradish Peroxidase (HRP)-conjugated antibody specific for ORM1 / alpha 1 Acid Glycoprotein is added to each well and incubate. After washing away any unbound antibody-enzyme reagent, a substrate solution (TMB) is added to the wells and color develops in proportion to the amount of ORM1 / alpha 1 Acid Glycoprotein bound in the initial step. The color development is stopped by the addition of acid and the intensity of the color is measured at a wavelength of 450nm  $\pm$ 2nm. The concentration of ORM1 / alpha 1 Acid Glycoprotein in the sample is then determined by comparing the O.D of samples to the standard curve.

## **MATERIALS PROVIDED & STORAGE INFORMATION**

Store the unopened kit at 2-8°C. Use the kit before expiration date.

<b>NO</b>	<b>Component</b>	<b>Quantity</b>	<b>Storage information</b>
C1	Antibody-coated microplate	8 X 12 strips	4°C
C2	Standard (Lyophilized)	1 vial	4°C
C3	5X Diluent buffer	50 ml	4°C
C4	HRP-conjugate Antibody	150 µl	4°C
C5	20X Wash buffer	50 ml	4°C
C6	TMB substrate	12 ml (ready to use)	4°C (Protect from light)
C7	STOP solution	12 ml (ready to use)	4°C
C8	Plate sealer	3 strips	Room temperature

## **MATERIALS REQUIRED BUT NOT PROVIDED**

- Microplate reader capable of measuring absorbance at 450nm
- Pipettes and pipette tips
- Deionized or distilled water
- 37°C oven or incubator
- Automated microplate washer (optional)

## **TECHNICAL HINTS AND PRECAUTIONS**

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid samples.
- Upon received, store all component in the kit at 2-8°C at all times.
- If crystals are observed in the 20X Wash buffer, warm to RT (not more

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than 50°C) until the crystals are completely dissolved.

- Ensure complete reconstitution and dilution of reagents prior to use.
- All materials should be equilibrated to room temperature (RT, 22-25°C) 20 min before use.
- All reagents should be mixed by gentle inversion or swirling prior to use. Do not induce foaming.
- Before using the kit, spin tubes and bring down all components to the bottom of tubes.
- Mix the contents of the microplate wells thoroughly by microplate shaker for 1 min or gently tap the plate to ensure good test results. Please mix carefully to avoid well-to-well contamination. Do not reuse microwells.
- The TMB Color developing agent should be colorless and transparent before using.
- Use reservoirs only for single reagents. This especially applies to the substrate reservoirs. Using a reservoir for dispensing a substrate solution that had previously been used for the conjugate solution may turn solution colored. Do not pour reagents back into vials as reagent contamination may occur.
- Do not let wells dry during assay; add reagents immediately after completing the rinsing steps.
- Avoid using reagents from different batches.
- It is highly recommended that the standards, samples and controls be assayed in duplicates.
- Change pipette tips between the addition of different reagent or samples.

## **SAMPLE COLLECTION & STORAGE INFORMATION**

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

**Serum**- Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Collect serum and assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Urine**- Collect the urine by micturating directly into a sterile container. Remove impurities by centrifugation at 10,000 x g for 1 min. Collect the supernatants and assay immediately or aliquot and store samples at  $-20^{\circ}\text{C}$  up to 1 month or  $-80^{\circ}\text{C}$  up to 6 months.

Note:

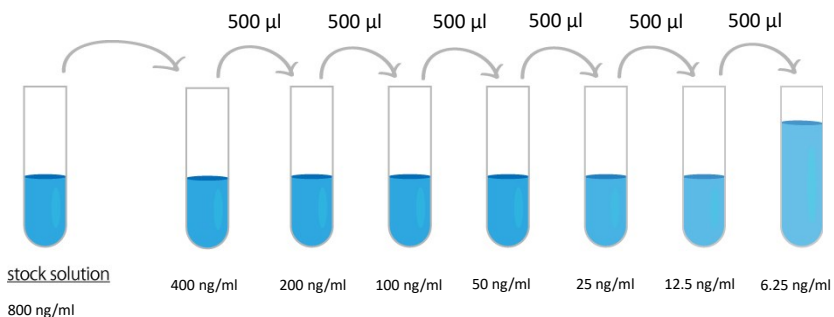
- a) Do not use haemolytic, icteric or lipaemic specimens.
- b) Samples containing sodium azide should not be used in the assay.

## **REAGENT PREPARATION**

- **1X Wash buffer:** Dilute 20X Wash buffer into distilled water to yield 1X Wash buffer.  
\*The diluted Wash buffer is stable for 4 weeks at 2°C to 8°C.
- **1X Diluent Buffer:** Dilute 5X Diluent Buffer into distilled water to yield 1X Diluent Buffer.
- **1X HRP-conjugate Antibody:** 20 minutes before use, dilute **100X HRP-conjugate Antibody** into 1X Diluent Buffer to yield **1X HRP-conjugate Antibody**.
- **Sample:** If the initial assay found samples contain ORM1 / alpha 1 Acid Glycoprotein higher than the highest standard, the samples can be diluted with diluent buffer and then re-assay the samples. For the calculation of the concentrations this dilution factor has to be taken into account.  
**(It is recommended to do pre-test to determine the suitable dilution factor).**
- **Standards:** Reconstitute the standard with **1 ml 1X diluent buffer** to yield a stock concentration of 800 ng/ml. Keep the buffer in the vial for at least **15 min at RT** to make sure the standard is dissolved completely before making serial dilutions. The 1X diluent buffer serves as zero standard (0 ng/ml), and the rest of the standard serial dilution can be diluted as according to the suggested concentration below: **400 ng/ml, 200 ng/ml, 100 ng/ml, 50 ng/ml, 25 ng/ml, 12.5 ng/ml, 6.25 ng/ml**. DO NOT reuse the reconstituted standard.

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Dilute ORM1 / alpha 1 Acid Glycoprotein standard as according to the table below:

Standard	ORM1 / alpha 1 Acid Glycoprotein Conc.	µl of diluent	µl of standard
S7	400 ng/ml	500	500 (800 ng/ml Stock)
S6	200 ng/ml	500	500 (S7)
S5	100 ng/ml	500	500 (S6)
S4	50 ng/ml	500	500 (S5)
S3	25 ng/ml	500	500 (S4)
S2	12.5 ng/ml	500	500 (S3)
S1	6.25 ng/ml	500	500 (S2)
S0	0	500	0



## **ASSAY PROCEDURE**

**All materials should be equilibrated to room temperature (RT) 20 min before use.**

Standards, samples and controls should be assayed in duplicates.

1. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal it.
2. Add **100 µl** of standards, samples and zero controls (1X diluent buffer) into wells, gently tap the plate to mix well. Incubate for **60 minutes at RT**.
3. Aspirate each well and wash, repeating the process three times for a total **4 washes**. Wash by filling each well with 1x Wash Buffer (300 µl) using a squirt bottle, manifold dispenser, or autowasher, keep the Wash Buffer in the wells for 30 sec before remove. Complete removal of liquid at each is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating, decanting or blotting against clean paper towels.
4. Add **100 µl** 1X HRP-conjugate Antibody into each well, gently tap the plate to mix well. Cover wells and incubate for **10 minutes at RT**.
5. Aspirate each well and **wash as step 3**.
6. Add **100 µl** of TMB Reagent to each well, gently tap the plate to mix well. Incubate for **5-15 minutes at RT** in dark.
7. Add **100 µl** of Stop Solution to each well, gently tap the plate to mix well. The color of the solution should change from blue to yellow.
8. Read the OD with a microplate reader at **450 nm immediately**. It is recommended read the absorbance within 3 min after adding STOP solution.

## **CALCULATION OF RESULTS**

1. Calculate the average absorbance values for each set of standards, controls and patient samples.
2. Using linear graph paper, construct a standard curve by plotting the mean absorbance obtained from each standard against its concentration with absorbance value on the vertical (Y) axis and concentration on the horizontal (X) axis.
3. Using the mean absorbance value for each sample determine the corresponding concentration from the standard curve.
4. Automated method: The results in the IFU have been calculated automatically using a 4 PL (4 Parameter Logistics) curve fit. 4 Parameter Logistics is the preferred method. Other data reduction functions may give slightly different results.
5. arigo provides GainData®, an in-house development ELISA data calculator, for ELISA data result analysis. Please refer our GainData® website for details. (<https://www.arigobio.com/elisa-analysis>)
6. If the samples have been diluted, the concentration read from the standard curve must be further converted by the appropriate dilution factor according to the sample preparation procedure as described above.

## **QUALITY ASSURANCE**

### **Sensitivity**

The minimum detectable dose (MDD) of Rat orosomuroid 1 ranged from 12.5 - 400 ng/ml. The mean MDD was 6.25 ng/ml.

### **Specificity**

This assay recognizes natural and recombinant Rat orosomuroid 1. No significant cross-reactivity or interference with the factors below was observed:

### **Intra-assay and Inter-assay precision**

The CV values of both intra and inter precision fall below 10%.