

Phytic acid Assay Kit

ARG83416 Phytic acid Assay Kit can be used to measure Phytic acid in tissue extracts, cell lysate, cell culture media and other biological fluids.

Catalog number: ARG83416

Package: 96 wells

For research use only. Not for use in diagnostic procedures.

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INTRODUCTION

The (myo) phytate anion is a colorless species that has significant nutritional role as the principal storage form of phosphorus in many plant tissues, especially bran and seeds. It is also present in many legumes, cereals, and grains. Phytic acid and phytate have a strong binding affinity to the dietary minerals, calcium, iron, and zinc, inhibiting their absorption in the small intestine.

PRINCIPLE OF THE ASSAY

The Phytic acid Assay Kit can measure Phytic acid in tissue extracts, cell lysate, cell culture media and other biological fluids. The increase in absorbance at 660 nm is directly proportional to reactants of the reaction between substrate and phytic acid.

MATERIALS PROVIDED & STORAGE INFORMATION

Component	Quantity	Storage	
Microplate	1 X 96-well plate		
Standard	1 vial (lyophilized)	4°C	
Enzyme	1 vial (lyophilized)	-20°C	
Reaction Buffer	8 ml (ready to use)	4°C	
Reagent Dye I	1 vial (lyophilized)	4°C	
Reagent Dye II	1 vial (lyophilized)	4°C	
Reagent Dye III	10 ml	4°C	

MATERIALS REQUIRED BUT NOT PROVIDED

- Microplate reader capable of measuring absorbance at 660 nm
- Pipettes and pipette tips
- Deionized or distilled water

TECHNICAL HINTS AND PRECAUTIONS

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid samples.
- Store Enzyme at -20°C, all other components store at 4°C.
- Briefly spin down the reagents before use.
- It is highly recommended that the standards and samples be assayed in at least duplicates.
- Change pipette tips between the addition of different reagent or samples.

SAMPLE COLLECTION & STORAGE INFORMATION

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

<u>Cell and bacteria</u> - Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml distilled water for 5×106 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

<u>Tissue</u> - Weigh out 0.1 g tissue, homogenize with 1 ml distilled water on ice, centrifuged at 8000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

^{*}Note: liquid samples can detect directly.

REAGENT PREPARATION

- Standard: Add 1.25 ml of distilled water to dissolve standard. The concentration will be 4 mmol/L. Perform 2-fold serial dilution of the top standards to make the standard curve.
- Enzyme: Add 1.1 ml of distilled water to dissolve Enzyme stored at -20°C.
- Reagent Dye: Add 5 ml Dye Reagent III into Dye Reagent I and 1 ml Dye Reagent III into Dye Reagent III respectively to dissolve. Transfer all Dye Reagent II into Dye Reagent III, mix; then transfer all Dye Reagent I into Dye Reagent III (Must follow this step). The mixed Dye Reagent may store at 4 °C for 2-3 days. This solution should be prepared before use.
 - *Note: It should be yellow. If colorless, the solution is failure. If blue, the solution is polluted.

ASSAY PROCEDURE

Standards and samples should be assayed in at least duplicates.

- 1. <u>Sample wells:</u> Add **10 μl Sample** into Sample wells.
- 2. Standard wells: Add **10 μl Standard** into Standard wells.
- 3. Add **80 µl Reaction Buffer** to each wells.
- 4. Add 10 µl Enzyme to each wells.
- 5. Mix well. Incubate at 55°C for 10 min
- 6. Add **100 μl Reagent Dye** to each wells.
- 7. Mix well. Read the OD at 660nm

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Reagent	Sample	Standard	Blank		
Sample	10 μΙ	-	-		
Standard	-	10 μΙ	-		
Distilled water	=		10 μΙ		
Reagent Buffer	80 μΙ	80 μΙ	80 μΙ		
Enzyme	10 μΙ	10 μΙ	10 μΙ		
Mix well. Incubate at 55°C for 10 min					
Reagent Dye	100 μΙ	100 μΙ	100 μΙ		
Mix well. Read the OD at 660nm					

CALCULATION OF RESULTS

1. Calculate the average absorbance values for each set of samples, standard and blank.

2. Calculation:

A. Definition:

C_{Standard}: the standard concentration, 4 µmol /ml;

C_{Protein}: the protein concentration, mg/ml;

W: the weight of sample, g;

 V_{Sample} : the volume of reaction sample, 10 μ l = 0.01 ml;

 $V_{standard}$: the volume of standard, 10 μ l = 0.1 ml;

 V_{assay} : the volume of Assay Buffer, 1000 μ l = 1 ml.

N: the quantity of cell or bacteria, $N \times 10^4$

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B. Formula:

a). According to the protein concentration of sample

```
\label{eq:Phytic acid (mmol/g) = (CStandard X Vstandard) X (ODSample - ODblank)] / [(ODStandard - ODBlank) X (CProtein X VSample)]} \\
```

b). According to the weight of sample

```
Phytic acid (\mumol/g) =  [(C_{Standard} \ X \ V_{Standard}) \ X \ (OD_{Sample} - OD_{blank})] \ / \ [(OD_{Standard} - OD_{blank}) \ X \ (W \ X \ V_{Sample} \ / \ V_{assay})]
```

c). According to the volume of sample

```
Phytic acid (µmol/g) =

[(Cstandard X Vstandard) X (ODsample – ODblank)] / [(ODstandard - ODblank) X Vassay)]

=4 X (ODsample - ODblank) / (ODstandard - ODblank)
```

d). According to the cell or bacteria

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Phytic acid (\mumol /10<sup>4</sup> cell) = 
[(Cstandard X Vstandard) X (ODsample - ODblank)] / [(ODstandard - ODblank) X (N X Vsample / VAssay)]
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=4 X (OD_{Sample} - OD_{blank}) / $[(OD_{Standard} - OD_{Blank}) X N]$

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3. Detection range:

The detection range is from 0.04 mg/ml - 4mg/ml.

4. If the samples have been diluted, the calculated activity must be further converted by the appropriate dilution factor according to the sample preparation procedure as described above.

EXAMPLE OF TYPICAL RESULT

The following data is for demonstration only and cannot be used in place of data generations at the time of assay. Please note this data is for demonstration only and serial diluted standards are not necessary for this kit.

