



# **Coxiella burnetii / Q Fever Phase 1 antibody ELISA Kit**

Enzyme immunoassay kit for the quantification Coxiella burnetii / Q Fever Phase 1 antibody in Animals serum and milk.

Catalog number: ARG83064

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For research use only. Not for use in diagnostic procedures.

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### **INTRODUCTION**

The reference standard test for the serologic diagnosis of acute Q fever is the indirect immunofluorescence antibody (IFA) using *C. burnetii* antigen, performed on paired serum samples to demonstrate a significant (fourfold or more) rise in antibody titers. The first sample should be taken as early in the disease as possible, preferably in the first week of symptoms, and the second sample should be taken 3 to 6 weeks later. In most cases of Q fever, the first IgG IFA titer is typically low, or “negative,” and the second typically shows a fourfold or greater increase in IgG antibody levels. A negative test during the first week of illness does not rule out Q fever as a cause of illness. There are two distinct antigenic phases (phase I and phase II) to which humans develop antibody responses. In acute infection, an antibody response to *C. burnetii* phase II antigen is predominant and is higher than antibody levels to phase I antigen; the reverse is true in chronic infection which is associated with a rising phase I IgG titer that may be higher than phase II IgG.

IgM antibodies usually rise at the same time as IgG, near the end of the first week of illness, and remain elevated for months or longer and therefore provide limited diagnostic value on their own. Furthermore, IgM antibodies are less specific than IgG antibodies and more likely to result in a false positive. For these reasons, physicians requesting IgM serologic titers should also request concurrent IgG titers.

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### **PRINCIPLE OF THE ASSAY**

This assay employs the qualitative determination enzyme immunoassay technique. An antigen specific for Q Fever IgG antibody has been pre-coated onto a microtiter plate. Standards or samples are pipetted into the wells and any antibody present is bound by the immobilized antigen. After washing away any unbound substances, a Horseradish Peroxidase (HRP) labeled Protein conjugate is added to each microplate well and incubated. This conjugate binds to antigen-antibody complexes. After washing away any unbound reagent, a substrate solution (TMB) is added to the wells and color develops in proportion to the amount of Q Fever IgG antibody bound in the initial step. The color development is stopped by the addition of acid and the intensity of the color is measured at a wavelength of 450 nm  $\pm$  2 nm.

### **MATERIALS PROVIDED & STORAGE INFORMATION**

Store the unopened kit at 2-8 °C. Use the kit before expiration date.

<b>Component</b>	<b>Quantity</b>	<b>Storage information</b>
Antigen-coated microplate	1 plate	4°C.
Control A (Negative Control)	2 ml (ready for use)	4°C
Control B (Cut-off Control)	3 ml (ready for use)	4°C
Control C (Positive Control)	2 ml (ready for use)	4°C
HRP-Streptavidin conjugate	20ml (ready for use)	4°C (Protect from light)
Sample Diluent Buffer	100 ml (ready for use)	4°C
20X Wash buffer	50 ml	4°C
TMB substrate	15 ml	4°C (Protect from light)
STOP solution	15 ml	4°C

### **MATERIALS REQUIRED BUT NOT PROVIDED**

- Microplate reader capable of measuring absorbance at 450nm
- Pipettes and pipette tips
- Deionized or distilled water
- Automated microplate washer (optional)

### **TECHNICAL HINTS AND PRECAUTIONS**

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid samples.
- Store the kit at 4°C at all times.
- Briefly spin down the antibody conjugate concentrate and HRP-Streptavidin concentrate before use.
- If crystals are observed in the 20X Wash buffer, warm to RT (not more than 50°C) until the crystals are completely dissolved.
- Ensure complete reconstitution and dilution of reagents prior to use.
- It is highly recommended that the standards, samples and controls be assayed in duplicates.
- Change pipette tips between the addition of different reagent or samples.
- Samples contain azide cannot be assayed.

## **SAMPLE COLLECTION & STORAGE INFORMATION**

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

**Serum** - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at  $\leq -20$  °C. Avoid repeated freeze-thaw cycles.

**Milk** – Whole milk can be use after centrifuged for 15 minutes at 2000 x g at 4°C. The sample should be taken from below the oil layer.

## **REAGENT PREPARATION**

- **1X Wash buffer:** Dilute 20X Wash buffer into distilled water to yield 1X Wash buffer. Diluted wash buffer is stable for 5 days at RT.
- **Serum Samples:** Samples have to be diluted 1+100 with sample diluent. (e.g. Dispense 10 $\mu$ l sample and 1ml Sample Diluent Buffer into tubes to obtain a 1+100 dilution and thoroughly mix with a Vortex.)
- **Milk Samples:** Samples have to be diluted 1+10 with sample diluent. (e.g. Dispense 100 $\mu$ l sample and 1ml Sample Diluent Buffer into tubes to obtain a 1+10 dilution and thoroughly mix with a Vortex.)

### ASSAY PROCEDURE

All materials should be equilibrated to room temperature (RT) before use. Standards, samples and controls should be assayed in duplicates.

1. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal it.
2. Add **100 µl** of control and prepared samples into wells.
3. Cover wells and incubate for **60 minutes** at 37°C.
4. Aspirate each well and wash, repeating the process 2 times for a total 3 washes. Wash by filling each well with 1× Wash Buffer (350 µl) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating, decanting or blotting against clean paper towels.
5. Add **100 µl** of HRP-Streptavidin conjugate into all well. Cover wells and incubate for **30 minutes** at RT in dark.
6. Aspirate each well and wash as step 4.
7. Add **100 µl** of TMB substrate into all well. Cover wells and incubate for **15 minutes** at RT in dark.
8. Add **100 µl** of Stop Solution to each well.
9. Read the OD with a microplate reader at **450/620 nm** immediately. It is recommended reading the absorbance within 30 min after adding Stop solution.

### MEASUREMENT

1. Adjust the ELISA microwell plate reader to zero using the Substrate Blank.  
If- due to technical reasons- the ELISA microwell plate reader cannot be adjusted to zero using the Substrate Blank, subtract its absorbance value from all other absorbance values measured in order to obtain reliable results.
2. Measure the absorbance of all wells at 450 nm and record the absorbance values for each standard/control and sample in the plate layout. Bichromatic measurement using a reference wavelength of 620 nm is recommended. Where applicable calculate the mean absorbance values of all duplicates.

### RESULTS

1. Run Validation Criteria

In order for an assay to be considered valid, the following criteria must be met:

- Substrate Blank: Absorbance value < 0.100
- Negative Control: Absorbance value < 0.200 and < Cut-off
- Cut-off Control: Absorbance value 0.150 – 1.300
- Positive Control: Absorbance value > Cut-off

If these criteria are not met, the test is not valid and must be repeated.

2. Calculation of Results

The Cut-off is the mean absorbance value of the Cut-off Control determinations. Example:

Absorbance value Cut-off Control (0.44) + absorbance value Cut-off



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control (0.42) =  $0.86 / 2 = 0.43$

Cut-off = 0.43

### 3. Results in Units [U]

[Units = U] = (Sample (mean) absorbance value x 10) / Cut-off

Example:  $(1.591 \times 10) / 0.43 = 37 \text{ U}$

### 4. Interpretation of Results

- Positive: > 11 U

Antibodies against the pathogen are present.

There has been a contact with the antigen (pathogen resp. vaccine).

- Equivocal: 9-11 U

Antibodies against the pathogen could not be detected clearly.

It is recommended to repeat the test with a fresh sample in 2 to 4 weeks. If the result is equivocal again the sample is judged as negative.

- Negative: < 9 U

The sample contains no antibodies against the pathogen.

A previous contact with the antigen (pathogen resp. vaccine) is unlikely.

Diagnosis of an infectious disease should not be established on the basis of a single test result. A precise diagnosis should take into consideration clinical history, symptomatology as well as serological data.

In immunocompromised patients and newborns serological data only have restricted value.